

LACTOSE INTOLERANCE: GENETICS OF LACTASE POLYMORPHISMS, DIAGNOSIS AND NOVEL THERAPY

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*Lactose intolerance is a common disorder affecting an individual's ability to digest lactose present in milk or any food product. Lactose intolerance is caused by the deficiency of β -galactosidase (lactase) in the digestive tract. Diagnosis of lactose intolerance is not so simple and straightforward clinically. Many biochemical and genetic tests have been developed for the determination of lactose intolerance. Several case reports indicate wherein subjects have self-diagnosed being lactose intolerant. There is an emerging link of this disorder with human gene polymorphism, where genetic basis has been used as a diagnostic tool. The high prevalence of this condition among children and adults has compelled the production of lactose-free foods. Additionally, external enzyme supplementation has been looked at as an alternative protective mechanism in lactose intolerant subjects. This review highlights the genetic variants of lactase polymorphism and theranostic (therapeutic and diagnostic) strategies for lactose intolerance. **Biomed Rev 2014; 25: 35-44***

Keywords: Lactose intolerance, lactase (β -galactosidase) deficiency and persistence, LCT- gene polymorphism, biochemical and genetic tests, therapy

INTRODUCTION

Lactose is a disaccharide present in mammalian milk and is essential for the nourishment of newborn infants. It is hydrolyzed by the intestinal brush-border enzyme, lactase, into absorbable sugars, namely glucose and galactose to provide energy. In most infants, intestinal lactase activity is maximal during the postnatal period. However, after 2–12 years of age, there oc-

curs a segregation into two distinct groups, viz, “lactase non-persistence group” with low lactase activity (hypolactasia) and a “lactase-persistence group” of individuals who retain their neonatal level of lactase activity even in adulthood (1-3).

Lactose intolerance is caused by the deficiency of β -galactosidase (lactase) in the digestive tract. The typical

Received 11 November 2014, revised 20 November 2014, accepted 1 December 2014

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clinical symptoms consist of abdominal pain and distension of gastrointestinal tract, borborygmi, and flatulence. Sometimes diarrhoea may occur from 30 minutes and 2 hours after the ingestion of lactose (4). Lactase deficiency results in lack of absorption of non-digested lactose, which causes luminal water retention and leads to iso-osmolarity of chyme. Consequently, excessive fluid retention causes abdominal pain, nausea, and diarrhoea. Bacterial fermentation of lactose in the distal small intestine and colon further aggravate these symptoms.

Because of the acute diarrhoeal condition that occurs after drinking milk, people who suffer from this disorder generally avoid a lactose-containing diet such as milk and milk products (5). Nevertheless, milk is calcium and nutrient-rich food and an important part of a healthy diet (6). Avoidance of milk during childhood is a significant risk factor for retarded growth and development as well as low bone density (7). Those who avoid milk, due to lactose intolerance, consume significantly less calcium and suffer from poorer health and bone formation, and higher risk of osteoporosis (7). Some studies have suggested that the prevalence of lactose intoler-

ance is a global issue. It is estimated that the worldwide population incidence of this condition is around 30.5% in children of 11–13 years (8), and about 55.1% in Chinese adults (9). Also, high incidence of lactase deficiency has been reported among Hispanic people (50% to 80%), African-American and Ashkenazi Jewish population (60% to 80%), and is almost 100% in Asian and American Indian people (10).

As summarized in Table 1, lactose intolerance is observed in various distinct forms such as congenital, primary and secondary. Congenital lactase deficiency is associated with the least lactase activity. Congenital hypolactasia is a single autosomal recessive disorder and is an extremely rare condition (11). Primary adult-type hypolactasia is an autosomal recessive condition, resulting from the physiological decline of lactase enzyme activity in the intestinal cells, and occurs in a large proportion of individuals. A single nucleotide polymorphism, C/T-13910, 14 kb upstream the lactase gene, has recently been correlated with lactase persistence/non persistence in several populations (12, 13). Secondary causes of hypolactasia, such as celiac disease, gastroenteritis and Crohn's disease, may lead to transient lactase deficiency and

Table 1. Types of lactase deficiencies and their associated risk factors (References cited are shown in brackets)

Type of lactase deficiency	Risk factors / causes/triggers
Congenital lactase deficiency	This condition has been diagnosed in less than 50 people world-wide (15). It becomes apparent at birth with persistent diarrhoea soon after milk is introduced. These children otherwise have a normal intestinal mucosa (16).
Primary lactase deficiency	It occurs when there is a gradual reduction in lactase production. It may not become clinically evident until late adolescence (17). It's prevalent in those geographical groups where the ancestors did not drink milk as a nutrient (15, 18).
Secondary lactase deficiency	It occurs as a result of gastrointestinal illness that alters the nature of the gut mucosa (19). Cryptosporidiosis, giardiasis and other parasitic infections lead to lactose malabsorption; it is very common in the children with rotaviral diarrhoea (20). It may occur in association with celiac disease, Crohn's disease and HIV (21, 22). Secondary lactase deficiency may also occur due to certain drugs like tetracycline and methotrexate that causes villous atrophy. Alcohol may initiate or worsen the lactose intolerance (18).
Developmental lactase deficiency	Premature infants may experience this deficiency. However this condition is temporary and improves as the intestinal mucosa matures (23).

appearance of adverse abdominal symptoms (14).

The present review highlights the genetic variants of lactase polymorphism and theranostic (therapeutic and diagnostic) strategies for lactose intolerance.

BIOCHEMISTRY AND GENETICS OF LACTASE

Lactase is located on the apical surface (glycocalyx) of brush border enterocytes where it is anchored into the membrane by its C-terminal, with the bulk of the molecule projecting into the lumen of the gut (24). It is a large glycoprotein, with two active sites, that can catalyze the hydrolysis of a variety of β -glucosides, such as phlorizin, flavonoid glucosides (25), and pyridoxine-5'- β , -D glucoside (26), and β -galactosides in addition to lactose. Lactase is encoded by a single gene (*LCT*) which is approximately 50 kbp located on chromosome 2 (27, 28). The gene has 17 exons (27) and encodes an mRNA transcript for a preproprotein of 1927 aminoacid residues (29). This is composed of a putative signal peptide of 19 aminoacid residues, a large pro-portion of 849 amino acids and a mature protein that contains two catalytic sites, and at the C-terminal, a membrane-spanning domain and short cytoplasmic domain. *LCT* shows a four-fold internal homology, which suggests that it arose by two duplication events (24). Pro-lactase is proteolytically processed to a smaller protein (30-32) and two of the four homologous regions occur in the cleaved pro-portion of the molecule, which does not have a catalytic function, but probably has a chaperone function, in that it seems to play a role in transporting the molecule to the cell surface (33-38). There is one active site in each of the domains of the mature protein. Although details have been disputed by some investigators, it is now considered that the active site at Glu1273 in domain III is responsible for hydrolysis of glucosides such as phlorizin, whereas the other in domain IV, at Glu1749, catalyzes the hydrolysis of galactosides such as lactose (39, 40).

It has been reported that lactase is O-glycosylated through serines and threonines as well as N-glycosylated (through asparagine), and this glycosylation probably affects enzymatic activity as well as folding and intracellular transport (41). Lactase expression is restricted to the enterocytes of small intestine, at highest level being in the mid-jejunum (42).

LACTOSE INTOLERANCE SYMPTOMATOLOGY

Gut related symptoms

Hypolactasia may not cause any discomfort unless lactose-containing food is consumed. Colonic microflora ferment un-

digested lactose in the intestinal lumen, which leads to production of short-chain fatty acids, hydrogen, carbon dioxide, and methane as by-products causing flatulence, bloating and abdominal pain. Undigested lactose acidifies the colon and increases the osmotic load, resulting in formation of foamy, voluminous and aqueous stools (43). However, some patients can experience constipation due to decreased intestinal motility, possibly caused by production of methane (43).

Other symptoms

The clinical presentation of lactose intolerance is not just restricted to gut symptoms. Other complaints such as headache, vertigo, memory impairment, lethargy, muscle and joint pains, allergy, cardiac arrhythmia, mouth ulcers, and sore throat have been reported (4, 44). Colonic bacteria generate toxic metabolites by lactose fermentation such as acetaldehyde, ethanol, acetone, peptide and protein toxins, which can alter many cell signalling mechanisms and are possibly responsible for these symptoms (44).

There is a considerable individual variability in the severity of symptoms, depending upon the amount of lactose ingested and the patient's ability to digest it. Factors contributing to this variability include osmolarity and the fat content of lactose-containing food, ability of colonic microflora to ferment lactose, gastric emptying rate, colonic water absorption capacity, intestinal transit time and individual perception of abdominal pain and discomfort (43, 45).

EMERGING LINKS WITH GENETICS: LACTASE PERSISTENT ALLELES AND LACTASE POLYMORPHISMS

As indicated above, the *LCT* gene is 49.3 kbp in length and is located on the long (q) arm of chromosome 2 at position 21. It contains 17 exons and is translated into a 6 kb transcript (27-29). Individuals with hypolactasia and lactase persistence have identical coding sequences, except for a few silent mutations; hence, both lactases are identical (46). Two variants were found to be associated with lactase persistence: A polymorph variant, *LCT*-13910C.T, in intron 13 of the *MCM6* gene that is 13 910 bp from the initiation codon of *LCT*, while the *LCT*-22018G, a variant in intron 9 of *MCM6* gene upstream of the *LCT* locus 22,018 bp was associated (1, 2, 47). This association was confirmed in a study of DNA collected from subjects of Finnish, South Korean, Italian, German, French, or Caucasian or African-North-American descent (1,47). Both genotypes of *LCT*-13910CT and *LCT*-13910TT were associated with the lactase-persistence pheno-

type, indicating that the presence of one single lactase-persistence allele in the heterozygous state has a dominant effect, rendering the person a lactose digester (48).

PROMOTOR SEQUENCE STUDIES

Functional *in vitro* studies of these polymorphic alleles have shown that *LCT*-13910T (1, 49, 50), *LCT*-13907G, *LCT*-13915G, and *LCT*-14010C act as enhancers of the *LCT* promoter (51). These effects are most likely mediated by the Oct-1 transcriptional factor binding site in the variant enhancer and by HNF1 α binding in the *LCT* promoter (49, 52). *LCT* gene regulation of lactase-persistence alleles occurs at the transcriptional level. *LCT* mRNA levels, which are distinguished by polymorphic markers in the coding region of *LCT*, were several times higher in individuals with *LCT*-13910T/-22018A alleles than in individuals with *LCT*-13910C/-22018G alleles (1). It has also been generally agreed that nonpersistent individuals have lower levels of lactase mRNA [53-55].

Numerous transcription factors (Cdx2, GATA-4, GATA-5, GATA-6, and HNF1 α) activate the *LCT* promoter in intestinal cell culture at the -100 to -20 bp binding site regions of *LCT* which are repressed by PDX-1 (1). Mutation of the PDX-1 binding site does not prevent *LCT* promoter repression, which suggests that PDX-1 might function by binding to another DNA binding site or by inhibiting other transcriptional factors. PDX-1 over expression resulted in strong repression of Cdx2 and HNF1 α activation of the *LCT* promoter (1). However, the exact mechanism for down regulation of *LCT* after weaning still remains unknown.

CONGENITAL LACTASE DEFICIENCY

While secondary loss of lactase in children is a frequent problem, resulting from viral infection and allergy (56), true congenital deficiency of lactase is very rare indeed. Congenital lactase deficiency is one of the severe gastrointestinal disorders characterized by watery diarrhoea shortly after the first feed with breast milk or lactose containing formulas (57). It is a rare autosomal disorder which occurs due to mutations in the coding regions of lactase, *LCT* gene. The *LCT* gene consists of 17 exons encoding 1927 amino acids comprising four homologous domains I-IV. Domain IV harbours the lactase activity mutation in exon 9 and is responsible for the truncation of lactase. One such case was recently reported in a Japanese female infant who had two mutations in the *LCT* gene in a heterozygous form: c.4419C> G (p.Y1473X)

in exon 10 and c.5387 delA (p.D1796fs) in exon 16, these mutations occurred in domain IV and was considered causative for congenital lactase deficiency (58).

FALLACIES FOR SELF-DIAGNOSIS OF LACTOSE INTOLERANCE

Due to an increased public awareness about the prevalence of lactose intolerance, there have also been several misunderstandings about self-diagnosis (59). In addition, there is also a little understanding that lactose *maldigestion* (incomplete lactose digestion due to low levels of lactase) is not synonymous with lactose *intolerance* (symptoms such as bloating, cramps and diarrhoea that may or may not occur in association with undigested lactose in the intestinal tract). Many studies have identified that the majority of those with lactose maldigestion do not experience the symptoms of lactose intolerance after consuming moderate quantities of lactose (60-70), and consequently avoid the lactose containing foods. One such food is milk, which is the richest source of calcium, vitamins, fatty acids, and low intake of milk can lead to serious health complications, including chronic disease like osteoporosis (59). On the contrary, it has also been reported that consuming dairy foods may actually contribute to improve the tolerance to lactose. Thus, individuals with self-reported lactose intolerance are unlikely to meet the appropriate levels for calcium from food sources alone. Dietitians and other health professionals can play an important role in informing these individuals about how to include lactose-containing foods without experiencing adverse symptoms, as well as providing information on other food sources of calcium, and further if needed, advice on appropriate supplementations.

IMPLICATIONS OF LACTOSE INTOLERANCE AND OTHER DISEASES

In addition to being a major cause of inconvenience in itself, lactose intolerance could be associated with other health disorders like cystic fibrosis, an autosomal recessive disorder characterized by the loss of function of the cystic fibrosis transmembrane conductance regulator (CFTR) (71, 72). This occurs due to more than 1700 different mutations in the *CFTR* gene, the most common of which is the loss of phenylalanine at the 508 position of the CFTR protein resulting in its misfolding, a faulty posttranslational processing and endoplasmic reticulum regulated degradation (73). Reduced bone mineral density is a common malady affecting cystic fibrosis patients. Since, meat cannot by itself serve as a nutritional source (due to absence of pancreatic elastase-1 in

pancreatic supplements, an enzyme required to digest elastin fibres in meat) for exocrine pancreatic insufficient cystic fibrosis patients, the only alternative is dairy products (71). Although not typical for cystic fibrosis, lactose intolerance can hamper dairy product consumption and fail to help cystic fibrosis patients to increase their bone mineral density in severe cases.

Lactose intolerance could also be associated with epilepsy, which affects about 45 million people across the globe (74-76). Neural cells contain glycosphingolipids whose biosynthesis requires galactose which is derived from lactose and other carbohydrates. Lactose intolerance impairs the production of galactose which could lead to malfunctioning of neurons resulting in epileptic episodes (77).

DIAGNOSIS OF LACTOSE INTOLERANCE

Previously, the most reliable method available for detecting lactose intolerance was a direct biochemical assay of lactase activity from a jejunal sample which was performed with a glucose oxidase reagent, that detects glucose liberated from lactose, with a cut off value of 10 U/g protein (1,2). However due to the invasiveness of jejunal biopsy, this method was then replaced by endoscopic duodenal biopsy (78,79).

Lactose tolerance tests have been developed to confirm the ability of intestinal lactase to hydrolyze lactose so as to avoid intestinal biopsies. In this technique, blood glucose levels are measured before and after an oral load of lactose at pre-specified time intervals, with a maximum rise of 20 mg/dL, indicating lactose tolerance (80). Oral ethanol administration before lactose load is used to inhibit galactose metabolism for the determination of the blood maximum rise of glucose (at least 20 mg/dL) and galactose (at least 10 mg/dL), thereby indicating lactose tolerance. Thus, galactose concentration in combination with glucose concentration improves the correlation with jejunal lactase activity than using only glucose maximum rise after lactose load (81).

Despite of all the indirect lactose tolerance tests currently available, breath hydrogen concentration after ingestion of 50 g of lactose was considered the most suitable test for population screening for lactase deficiency (82). However, the use of the 50 g lactose dose has been criticized, because it is equivalent to 4–5 cups of milk, an amount that is ideally far more than an individual can usually ingest at one time, so an oral load of 25 g may be considered a more appropriate amount, with high sensitivity and specificity (80,83).

Interestingly, the discovery of lactase-persistence al-

leles resulted in the advent of genetic tests for diagnosis of lactase non-persistence by polymerase chain reaction restriction fragment length polymorphism (84-86), real-time polymerase chain reaction [87-89], and pyrosequencing technology (90). Compared with the lactose hydrogen breath test, the genetic test has numerous advantages such as it is simple, non-invasive, and more comfortable examination that does not provoke symptoms of lactose intolerance and is less cumbersome (85). However, other polymorphic variants in Europeans (*LCT*-13914G.A)50 and in African and Arab populations (*LCT*-13907C.G, *LCT*-13913T.C, and *LCT*-13915T.G, close to *LCT*-13910C.T.) affect the diagnostic accuracy of *LCT*-13910C.T typing by altering the melting profiles of the real-time polymerase chain reaction kit [89]. The reverse-hybridization strip assay based on multiplex DNA amplification and ready-to-use membrane test strips that detect *LCT* polymorphic variants (-13907C.G, -13910C.T, -13913T.C, -13914G.A, -13915T.G, and -22018G.A) and inevitably represents a reliable tool for genetic diagnosis of lactase non-persistence helps in overcoming the interference of different melting profiles of the real-time polymerase chain reaction kit by the other polymorphic variants (91). The genetic test provides a more direct result, i.e., a hypolactasia or lactase persistence genotype, whereas interpretation of the lactose breath test depends on several variants such as the cut off level, dose of lactose given, and duration of the test and age of the individual, and is also expensive.

NOVEL TREATMENT INTERVENTIONS FOR MANAGEMENT OF LACTOSE INTOLERANCE

The initial recommendation for management of lactose intolerance is to aim for remission of symptoms through temporary avoidance of milk and dairy products. Most individuals with lactose malabsorption can tolerate up to 12 g of lactose without significant symptoms. After the initially restricted diet, lactose should be gradually reintroduced until the patient's threshold for symptoms is reached (92). The main pharmacological measures include lactase supplements, lactose-hydrolyzed or lactose-reduced milk, probiotics (93), and colonic adaptation.

NON-LACTOSE INFANT FORMULAS

Infants who cannot tolerate any lactose may only be treated by excluding lactose from diet. This could be achieved by using lactose-free infant formulas or incubating feeds with the enzyme lactase which can break the sugar into its component

parts. In case of primary lactose intolerance where the degree of lactase deficiency varies, the use of lactose-free formula may help to relieve symptoms. In lactose-free formula, the carbohydrate source is glucose rather than lactose, therefore these milk products have a greater potential to cause dental caries. This is because lactose is a non-cariogenic sugar whereas glucose is cariogenic (94).

Soy-based infant formulas have been often preferred to lactose-free formula. However, they may not be safe because contain phytoestrogens; note, soy formulas are no more in usage for infants. A recent report suggests that phytoestrogens in soy infant formulas are capable of inhibiting the action of an enzyme involved in iodination of thyroxine (thyroid peroxidase, TPO) through competitive inhibition (95). It has not yet clearly established that the levels of free phytoestrogen in infants' plasma are sufficient to significantly inhibit TPO. Hence, the clinical significance of phytoestrogen consumption in the presence of adequate iodine intake still remains unclear.

ENZYME THERAPY: ENDOGENOUS β -GALACTOSIDASE TO ALLEVIATE LACTOSE INTOLERANCE

In addition to decreasing the lactose concentration in milk products, there have been several techniques that have been used to produce low-lactose milk, such as β -galactosidase enzymes hydrolysis techniques or combinative techniques with ultrafiltration and enzymes hydrolysis (96, 97). However, they have also been known to alter the quality of the milk products and their commercial value (98). Enhancing the intestinal β -galactosidase activity of lactose intolerant subjects has gained more importance. Exogenous β -galactosidase was usually prescribed for lactose intolerant subjects. However, most of the supplemental β -galactosidase displayed poor stability in human gut. Interestingly, the endogenous β -galactosidase expressed in intestinal microbes has been reported to help humans in lactose usage (99), and is one of the promising treatment strategies, since it is associated with the promotion of beneficial microorganism in the gut (100). Therefore, the enhancement of β -galactosidase in the intestinal microflora of humans together with selected probiotics may be a promising approach in lactose intolerance management. An evidence to this fact was provided by a recent *in vivo* study conducted to evaluate the alleviation of lactose intolerance symptoms in post-weaning Balb/c mice, which were orally administered with 1×10^6 CFU or 1×10^8 CFU of *L. lactis* MG1363/FGZW daily for 4 weeks before lactose challenge. It was observed that in comparison with naïve mice, the mice administered *L. lactis* MG1363/FGZW

showed significant reduction of diarrhea symptoms accompanied by lesser weight of total feces within 6 h post-challenge and suppressed intestinal motility after lactose challenge.

CONCLUSION

We have highlighted the genetic variants of lactase polymorphism and theranostic strategies for lactose intolerance. A significant progress has been made in our understanding of lactose intolerance. Management of lactose intolerance has improved over the years with the introduction of newer and better options which help to bypass the inevitable pitfalls of simple lactose avoidance. There is no single gold standard test available for the diagnosis of lactose intolerance. The lactose breath test, although considered the best method may be influenced by several factors. Genetic testing has been a new tool for the diagnosis of hypolactasia/lactase persistence, but may not detect all the single nucleotide polymorphisms associated with this disorder. It appears that up to 12 g of lactose is well tolerated by lactase non-persistence individuals, which may negate the need for restrictions on lactose-hydrolyzed milk, fermented and matured milk products, hence preventing any subsequent deleterious effects on bone mass density.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

REFERENCES

1. Troelsen JT. Adult-type hypolactasia and regulation of lactase expression. *Biochim Biophys Acta* 2005;1723:19–32.
2. Rasinperä H, Savilahti E, Enattah NS, Kuokkanen M, Tötterman N, Lindahl H, *et al.* A genetic test which can be used to diagnose adult-type hypolactasia in children. *Gut* 2004;53:1571–1576. DOI: 10.1136/gut.2004.040048.
3. Rasinperä H, Kuokkanen M, Kolho KL, Lindahl H, Enattah NS, Savilahti E. Transcriptional down regulation of the lactase (*LCT*) gene during childhood. *Gut*. 2005;54:1660–1661. DOI: 10.1136/gut.2005.077404.
4. Harrington LK, Mayberry JF. A re-appraisal of lactose intolerance. *Int J Clin Pract* 2008;62:1541–1546. DOI: 10.1111/j.1742-1241.2008.01834.x
5. He M, Yang YX. Lactase deficiency and lactose intolerance. *Foreign Med Sci-Hygiene* 1999;6:339–342.
6. Horner TW, Dunn ML, Eggett DL, Ogden LV.

- β -Galactosidase activity of commercial lactase samples in raw and pasteurized milk at refrigerated temperatures. *J Dairy Sci* 2011;94:3242–3249. DOI: 10.3168/jds.2010-3742.
7. Savaiano D. Lactose intolerance: an unnecessary risk for low bone density. *Nestle Nutr Workshop Ser Pediatr Program* 2011;67:161–171.
 8. Yang Y, He M, Cui H, Bian L. Study on the incidence of lactose intolerance of children in China. *Wei Sheng Yan Jiu* 1999;28:44–46.
 9. Xu Y, Wu XY, Lu YQ. Study on lactose intolerance. *Acad Period Farm Prod Process* 2008;11:89–91.
 10. Heyman MB, Committee on Nutrition. Lactose intolerance in infants, children, and adolescents. *Pediatrics* 2006;118:1279–1286
 11. Kuokkanen M, Kokkonen J, Enattah NS, Ylisaukko-Oja T, Komu H, Varilo T, *et al.* Mutations in the translated region of the lactase gene (LCT) underlie congenital lactase deficiency. *Am J Hum Genet* 2006; 78: 339-344
 12. Enattah NS, Sahi T, Savilahti E, Terwilliger JD, Peltonen L, Jarvela I. Identification of a variant associated with adulttype hypolactasia. *Nat Genet* 2002; 30: 233-237. DOI: 10.1038/ng826.
 13. Szilagyi A, Malolepszy P, Hamard E, Xue X, Hilzenrat N, Ponniah M, *et al.* Comparison of a realtime polymerase chain reaction assay for lactase genetic polymorphism with standard indirect tests for lactose maldigestion. *Clin Gastroenterol Hepatol* 2007; 5: 192-196.
 14. Paolo Usai-Satta, Mariella Scarpa, Francesco Oppia, Francesco Cabras. Lactose malabsorption and intolerance: What should be the best clinical management? *World J Gastrointest Pharmacol Ther* 2012; 3: 29-33. DOI: 10.4292/wjgpt.v3.i3.29.
 15. McBean LD, Miller GD. Allaying fears and fallacies about lactose intolerance. *J Am Diet Assoc* 1998; 98: 671-676.
 16. Bhatnagar S, Aggarwal R. Lactose intolerance. *BMJ* 2007; 334: 1331-1332.
 17. Rusnyk A, Still CD. Lactose intolerance. *J Am Osteopath Assoc* 2001; 101(4 Suppl Pt 1): S10-12.
 18. Lactose intolerance and milk allergy. Auckland Allergy Clinic. Available from <http://www.allergyclinic.co.nz/guides/21.html>.
 19. Swagerty DL Jr, Walling AD, Klein RM. Lactose intolerance. *Am Fam Physician* 2002; 65: 1845-1850.
 20. Heyman MB. Lactose intolerance in infants, children and adolescents. *Pediatrics* 2006; 118: 1279-1286. DOI: 10.1542/peds.2006-1721.
 21. Heaney RP. Ethnicity, bone status, and the calcium requirement. *Nutrition Research*. 2002; 22: 153-178
 22. Nicklas TA, Qu H, Hughes SO, He M, Wagner SE, Foushee HR *et al.* Self-perceived lactose intolerance results in lower intakes of calcium and dairy foods and is associated with hypertension and diabetes in adults. *Am J Clin Nutr* 2011; 94: 191-8. DOI: 10.3945/ajcn.110.009860.
 23. Westland S, Crawley H. *Specialised Infant Formula in the UK: Additional Information for Health Professionals*. First Steps Nutrition Trust. 2013.
 24. Dallas M Swallow. Genetics of lactase persistence and lactose intolerance. *Annu Rev Genet* 2003. 37:197–219. DOI: 10.1146/annurev.genet.37.110801.143820.
 25. Nemeth K, Plumb GW, Berrin JG, Juge N, Jacob R, Naim HY, *et al.* Deglycosylation by small intestinal epithelial cell betaglucosidases is a critical step in the absorption and metabolism of dietary flavonoid glycosides in humans. *Eur J Nutr* 2003; 42:29–42.
 26. Mackey AD, Henderson GN, Gregory JF 3rd. Enzymatic hydrolysis of pyridoxine-50-beta-D-glucoside is catalyzed by intestinal lactase-phlorizin hydrolase. *J Biol Chem* 2002;277:26858–64
 27. Boll W, Wagner P, Mantei N. Structure of the chromosomal gene and cDNAs coding for lactase-phlorizin hydrolase in humans with adult-type hypolactasia or persistence of lactase. *Am J Hum Genet* 1991;48:889–902.
 28. Harvey CB, Fox MF, Jeggo PA, Mantei N, Povey S, Swallow DM. Regional localization of the lactase-phlorizin hydrolase gene, LCT, to chromosome 2q21. *Ann Hum Genet* 1993;57:179–185.
 29. Mantei N, Villa M, Enzler T, Wacker H, Boll W, James P, *et al.* Complete primary structure of human and rabbit lactase-phlorizin hydrolase: implications for biosynthesis, membrane anchoring and evolution of the enzyme. *EMBO J* 1998; 7:2705–2713.
 30. Mesonero JE, Gloor SM, Semenza G. Processing of human intestinal prolactase to an intermediate form by furin or by a furin-like proprotein convertase. *J Biol Chem* 1998;273:29430–29436.
 31. Zecca L, Mesonero JE, GloorSM, Semenza G. Species differences in the sites of cleavage of pro-lactase to lactase supports lack of selective pressure. *Biochim Biophys Acta* 1999;1435:51–60.

32. Keller P, Zecca L, Boukamel R, Zwicker E, Gloor S, Semenza G. Furin, PC1/3, and/or PC6A process rabbit, but not human, prolactase-phlorizin hydrolase to the 180-kDa intermediate. *J Biol Chem* 1995;270:25722–25728.
33. Jacob R, Bulleid NJ, Naim HY. Folding of human intestinal lactase-phlorizin hydrolase. *J Biol Chem* 1995;270:18678–18684.
34. Jacob R, Peters K, Naim HY. The prosequence of human lactase-phlorizin hydrolase modulates the folding of the mature enzyme. *J Biol Chem* 2002;277:8217–8225.
35. Naim HY. The pro-region of human intestinal lactase-phlorizin hydrolase is enzymatically inactive towards lactose. *Biol Chem Hoppe-Seyler* 1995;376:255–258.
36. Naim HY, Jacob R, Naim H, Sambrook JF, Gething MJ. The pro region of human intestinal lactase-phlorizin hydrolase. *J. Biol. Chem.* 1994;269:26933–26943.
37. Oberholzer T, Mantei N, Semenza G. The pro sequence of lactase-phlorizin hydrolase is required for the enzyme to reach the plasma membrane. An intramolecular chaperone? *FEBS Lett* 1993;333:127–131.
38. Ouwendijk J, Peters WJ, van de Vorstenbosch RA, Ginsel LA, Naim HY, Franssen JA. Routing and processing of lactase-phlorizin hydrolase in transfected Caco-2 cells. *J. Biol. Chem.* 1998;273:6650–6655.
39. Arribas JC, Herrero AG, Martin-Lomas M, Canada FJ, He S, Withers, S.G. Differential mechanism-based labeling and unequivocal activity assignment of the two active sites of intestinal lactase/phlorizin hydrolase. *Eur. J. Biochem.* 2000;267: 6996.
40. Zecca L, Mesonero JE, Stutz A, Poiree JC, Giudicelli J, Cursio, R, *et al.* Intestinal lactase-phlorizin hydrolase (LPH): the two catalytic sites; the role of the pancreas in pro-LPH maturation. *FEBS Lett* 1998; 435:225–228.
41. Naim HY, Lentze MJ. Impact of O-glycosylation on the function of human intestinal lactase-phlorizin hydrolase. Characterization of glycoforms varying in enzyme activity and localization of O-glycoside addition. *J Biol Chem* 1992;267:25494–25504.
42. Newcomer A, McGill D. Distribution of disaccharidase activity in the small bowel of normal and lactase-deficient subjects. *Gastroenterology* 1966;51:481–488.
43. Lomer MC, Parkes GC, Sanderson JD. Review article: lactose intolerance in clinical practice – myths and realities. *Aliment Pharmacol Ther* 2008; 27: 93–103.
44. Matthews SB, Waud JP, Roberts AG, Campbell AK. Systemic lactose intolerance: a new perspective on an old problem. *Postgrad Med J* 2005; 81: 167–173. DOI: 10.1136/pgmj.2004.025551.
45. Suchy FJ, Brannon PM, Carpenter TO, Fernandez JR, Gilsanz V, Gould JB, *et al.* National Institutes of Health Consensus Development Conference: lactose intolerance and health. *Ann Intern Med* 2010; 152: 792–796.
46. Boll W, Wagner P, Mantei N. Structure of the chromosomal gene and cDNAs coding for lactase-phlorizin hydrolase in humans with adult-type hypolactasia or persistence of lactase. *Am J Hum Genet* 1991;48:889–890.
47. Enattah NS, Sahi T, Savilahti E, Terwilliger JD, Peltonen L, Javerla I. Identification of a variant associated with adult-type hypolactasia. *Nat Genet* 2002;30:233–237. DOI: 10.1038/ng826.
48. Mattar R, Ferraz de Campos Mazo D, Carrilho FJ. Lactose intolerance: diagnosis, genetic, and clinical factors. *Clin Exp Gastroenterol* 2012; 5: 113–121. DOI: 10.2147/CEG.S32368.
49. Lewinsky RH, Jensen TGK, Møller J, Stensballe A, Olsen J, Troelsen JT. T-13910 DNA variant associated with lactase persistence interacts with Oct-1 and stimulates lactase promoter activity in vitro. *Hum Mol Genet* 2005;14:3945–3953.
50. Enattah NS, Jensen TGK, Nielsen M, Lewinski R., Kuokkanen, M., Rasinpera, *et al.* Independent introduction of two lactase-persistence alleles into human populations reflects different history of adaptation to milk culture. *Am J Hum Genet* 2008;82:57–72.
51. Tishkoff SA, Reed FA, Ranciaro A, Voight BF, Babbitt CC, Silverman JS, *et al.* Convergent adaptation of human lactase persistence in Africa and Europe. *Nat Genet* 2007; 39:31–40.
52. Olds LC, Ahn JK, Sibley E. -13915*G DNA polymorphism associated with lactase persistence in Africa interacts with Oct-1. *Hum Genet* 2011;129:111–113.
53. Fajardo O, Naim HY, Lacey SW. The polymorphic expression of lactase in adults is regulated at the messenger RNA level. *Gastroenterology* 1994; 106:1233–41
54. Rossi M, Maiuri L, Fusco MI, Salvati VM, Fuccio A, *et al.* Lactase persistence versus decline in human adults: Multifactorial events are involved in downregulation after weaning. *Gastroenterology* 1997; 112:1506–1514.
55. Wang Y, Harvey C, Rousset M, Swallow DM. Expression of human intestinal mRNA transcripts during development: analysis by a semiquantitative RNA polymerase chain reaction method. *Pediatr Res* 1994;36:514–521.

56. Walker-Smith J. Diarrheal disease. *Nestle Nutr Workshop Ser* 1997;38.
57. Savilahti E, Launiala K, Kuitunen P. Congenital lactase deficiency. A clinical study on 16 patients. *Arch Dis Child* 1983;58: 246-252.
58. Uchida N, Sakamoto O, Irie M. Two novel mutations in the lactase gene in a Japanese infant with congenital lactase deficiency. *Tohoku J. Exp Med.*, 2012; 227: 69-72.
59. McBean LD, Miller GD. Allaying fears and fallacies of lactose intolerance. *J Am Dietetic Associ* 1998; 98: 671-676.
60. Rosado JL, Allen LH, Solomons NW. Milk consumption, symptom response, and lactose digestion in milk intolerance. *Am J Clin Nutr* 1987; 45:1457-1460.
61. Suarez FL, Savaiano DA, Levitt MD. A comparison of symptoms after the consumption of milk or lactose-hydrolyzed milk by people with self-reported severe lactose intolerance. *N Engl J Med* 1995;333:1-4.
62. Vesa TH, Korpela RA, Sahi T. Tolerance to small amounts of lactose in lactose maldigesters. *Am J Clin Nutr* 1996;64:197-201.
63. Suarez FL, Savaiano D, Arbisi P, Levitt MD: Tolerance to the daily ingestion of two cups of milk by individuals claiming lactose intolerance. *Am J Clin Nutr* 1997;65:1502-1506.
64. Hertzler SR, Clancy SM: Kefir improves lactose digestion and tolerance in adults with lactose maldigestion. *J Am Diet Assoc* 2003;103:582-587.
65. Jarvinen RM, Loukaskorpi M, Uusitupa MI: Tolerance of symptomatic lactose malabsorbers to lactose in milk chocolate. *Eur J Clin Nutr* 2003;57:701-705. DOI: 10.1038/sj.ejcn.1601600.
66. Scrimshaw NS, Murray EB: The acceptability of milk and milk products in populations with a high prevalence of lactose intolerance. *Am J Clin Nutr* 1998;48:1079-1159.
67. Vesa TH, Marteau P, Korpela R: Lactose intolerance. *J Am Coll Nutr* 2000;19:165S-175S.
68. Peuhkuri K, Vapaatalo H, Korpela R, Teuri U: Lactose intolerance - a confusing clinical diagnosis. *Am J Clin Nutr* 2000;71:600-602.
69. Buchowski MS, Semanya J, Johnson AO: Dietary calcium intake in lactose maldigesting intolerant and tolerant African-American women. *J Am Coll Nutr* 2002;21:47-54.
70. Suarez FL, Savaiano DA, Levitt MD: A comparison of symptoms after the consumption of milk or lactose-hydrolyzed milk by people with self-reported severe lactose intolerance. *N Engl J Med* 1995;333:1-4.
71. Madry E, Krasinska B, Drzymala-Czyz S, Sands D, Lisowska A, Grebowiec P, et al. Lactose malabsorption is a risk factor for decreased bone mineral density in pancreatic insufficient cystic fibrosis patients. *Eur J Hum Genet* 2012; 20: 1092-1095. DOI: 10.1038/ejhg.2012.52.
72. Madry E, Fidler E, Sobczynska-Tomaszewska A, Lisowska A, Krzyzanowska P, Pogorzelski A et al. Mild CFTR mutations and genetic predisposition to lactase persistence in cystic fibrosis. *Eur J Hum Genet* 2011; 19: 748-752. DOI: 10.1038/ejhg.2011.36.
73. Collawn JF, Fu L, Bebok Z. Targets for cystic fibrosis therapy: proteomic analysis and correction of mutant cystic fibrosis transmembrane conductance regulator. *Expert Rev Proteomics* 2010; 7: 495-506. DOI: 10.1586/epr.10.45.
74. Stein MA, Kanner AM. Management of newly diagnosed epilepsy: A practical guide to monotherapy. *Drugs* 2009; 69: 199-222. DOI: 10.2165/00003495-200969020-00005.
75. French JA, Pedley TA. Initial management of epilepsy. *N Engl J Med* 2008; 359: 166-76. DOI: 10.1056/NEJMc-p0801738.
76. Steinlein OK. Genetic mechanisms that underlie epilepsy. *Nat Rev Neurosci* 2004; 5: 400-408.
77. Yaman H, Karaoglu A, Cayci T, Akgul EO, Kurt YG, Tunc T et al. Epileptic seizures associated with lactose intolerance in a child: A causal relationship? *J Pediatr Neurol* 2012; 10: 151-54. DOI: 10.3233/JPN-2012-0551.
78. Lagman JM, Rowland R. Activity of duodenal disaccharidases in relation to normal and abnormal mucosal morphology. *J Clin Pathol* 1990;43:537-540.
79. Kuokkanen M, Myllyniemi M, Vauhkonen M, Helske T, Kääriäinen I, Karesvuori S, et al. A biopsy-based quick test in the diagnosis of duodenal hypolactasia in upper gastrointestinal endoscopy. *Endoscopy* 2006;38:708-712.
80. Law D, Conklin J, Pimentel M. Lactose intolerance and the role of the lactose breath test. *Am J Gastroenterol* 2010;105:1726-1728. DOI: 10.1038/ajg.2010.146.
81. Jussila J. Diagnosis of lactose malabsorption by the lactose tolerance test with peroral ethanol administration. *Scand J Gastroenterol* 1969;4:361-368.

82. Newcomer AD, McGill DB, Thomas PJ, Hofmann AF. Prospective comparison of indirect methods for detecting lactase deficiency. *N Engl J Med* 1975;293:1232–1236.
83. Romagnuolo J, Schiller D, Bailey RJ. Using breath tests wisely in a gastroenterology practice: an evidence-based review of indications and pitfalls in interpretation. *Am J Gastroenterol*. 2002;97:1113–1126.
84. Mattar R, Monteiro MS, Villares CA, Santos AF, Carrilho FJ. Single nucleotide polymorphism C/T-13910, located upstream of the lactase gene, associated with adult-type hypolactasia: validation for clinical practice. *Clin Biochem* 2008;41:628–630.
85. Büning C, Genschel J, Jurga J, Fiedler T, Voderholzer W, Fiedler EM, *et al*. Introducing genetic testing for adult-type hypolactasia. *Digestion* 2005;71:245–250.
86. Hogenauer C, Hammer HF, Mellitzer K, Renner W, Krejs GJ, Toplak H. Evaluation of a new DNA test compared with the lactose hydrogen breath test for the diagnosis of lactase non-persistence. *Eur J Gastroenterol Hepatol*. 2005;17:371–376.
87. Bodlaj G, Stöcher M, Hufnagl P, Hubmann R, Biesenbach G, Stekel H, *et al*. Genotyping of the lactase-phlorizin hydrolase –13910 polymorphism by lightCycler PCR and implications for the diagnosis of lactose intolerance. *Clin Chem* 2006;52:148–151.
88. Szilagyi A, Malolepszy P, Hamard E, Xue X, Hilzenrat N, Ponniah M, *et al*. Comparison of a real-time polymerase chain reaction assay for lactase genetic polymorphism with standard indirect tests for lactose maldigestion. *Clin Gastroenterol Hepatol* 2007;5:192–196.
89. Tag CG, Schiffers MC, Mohnen M, Gressner AM, Weiskirchen R. A novel proximal _13914G.A base replacement in the vicinity of the common-13910T/C lactase gene variation results in an atypical light cycler melting curve in testing with the MutaREAL lactase test. *Clin Chem* 2007;53:146–148.
90. Torbjörn K, Olsson LA. Simultaneous genotyping of the three lactose tolerance linked polymorphisms *LCT*-13907C.G, *LCT*-13910C.T and *LCT*-13915T.G with Pyrosequencing technology. *Clin Chem Lab Med* 2008;46:80–84.
91. Tag CG, Oberkanins C, Kriegshäuser G, Ingram CJ, Swallow DM, Gressner AM, *et al*. Evaluation of a novel reverse-hybridization StripAssay for typing DNA variants useful in diagnosis of adult-type hypolactasia. *Clin Chim Acta* 2008;392:58–62.
92. Montalto M, Curigliano V, Santoro L, Vastolo M, Cammarota G, Manna R *et al*. Management and treatment of lactose malabsorption. *World J Gastroenterol* 2006; 12: 187–191.
93. Malaguarnera G, Leggio F, Vacante M, Motta M, Giordano M, Biondi A *et al*. Probiotics in the gastrointestinal diseases of the elderly. *J Nutr Health Aging* 2012; 16(4): 402-410.
94. Heaney RP. Ethnicity, bone status, and the calcium requirement. *Nutrition Research*. 2002; 22: 153-178.
95. Tuohy P. *Soy based infant formula*. The Ministry of Health, Wellington, New Zealand. 1998.
96. Novalin S, Neuhaus W, Kulbe KD. A new innovative process to produce lactose-reduced skim milk. *J Biotechnol* 2005;119:212–218.
97. Chen CS, Hsu CK, Chiang BH. Optimization of the enzymic process for manufacturing low-lactose milk containing oligosaccharides. *Process Biochem* 2002;38:801–808.
98. Li W, Zhang XM, Lu Y. Lactose intolerance and the development of low-lactose milk. *China Dairy* 2004;2:35–37.
99. Masood MI, Qadir MI, Shirazi JH, Khan IU. Beneficial effects of lactic acid bacteria on human beings. *Crit Rev Microbiol* 2011;37:91–98.
100. Zhong Y. Probiotic and the study of the lactose intolerance. *Foreign Med Sci-Hygiene* 2003;2:101–105.
101. Li J, Zhang W, Wang C, Yu Q, Dai R, Pei X. Lactococcus lactis expressing food-grade β -galactosidase alleviates lactose intolerance symptoms in post-weaning Balb/c mice. *Appl Microbiol Biotechnol* 2012;96:1499–1506. DOI: 10.1007/s00253-012-3977-4.