

## NERVE GROWTH FACTOR, MAST CELLS AND ARTHRITIS

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### SUMMARY

• *Nerve growth factor is a well-characterized neurotrophic protein required for the survival and differentiation of a variety of neuronal cell types both in the peripheral and central nervous systems. Recent studies indicate that nerve growth factor also plays a role in cells originating in the immune system, since it is synthesized by cells of immune system lineage and its level increases during inflammatory responses. Moreover, it has been shown that cytokines such as interleukin-1 $\beta$  and tumor necrosis factor- $\alpha$  are potent inducers of nerve growth factor secretion. These studies were recently confirmed and extended by demonstrating that cells normally present in inflammatory tissues, such as mast cells and lymphocytes, express nerve growth factor receptors and are receptive to the action of nerve growth factor. The aim of the present review is to outline the current understanding of mast cells and nerve growth factor in autoimmune diseases and particularly in arthritis.*

### INTRODUCTION

• The term nerve growth factor (NGF) was introduced more than 40 years ago to define a newly discovered protein inducing neurite outgrowth from sensory and sympathetic ganglia *in vivo* and *in vitro* (1,2). NGF is synthesized and released in large amounts by the submaxillary salivary gland (SMG) of adult male mice (1-3). Lesser amounts have been found in snake venom (2), guinea pig prostate gland and seminal fluids of guinea pig and of bull (2,4), while nanogram quantities have been located in a variety of neuronal and non-neuronal cells, both in the central and peripheral nervous systems (2,5,6). The 2.5S NGF molecule isolated and purified

from the SMG is a dimer of two identical subunits linked together by non-covalent bonds and has a molecular weight of about 30 kD (7). The amino acid sequence and the primary structure of NGF from murine as well as from other species have been characterized and indicate highly conserved homology of murine NGF with other species, including humans (2,6,8-10).

The functional significance of NGF in the SMG and the physiological role of such a high concentration of NGF in these glands is largely unknown, although recent studies have shown that NGF released from the mouse SMG into the bloodstream following intraspecific aggressive behavior exerts an effect on at least two NGF target cells, the chromaffin cells and peritoneal mast cells (MC) (11-14).

In recent years, mainly through the use of exogenous NGF and NGF-antibody administration, it has been shown that NGF has specific cell targets not only in the peripheral and central nervous system, but also in the endocrine and immune systems. Accordingly, current knowledge indicates that NGF plays a functional role not only in neuropathological events associated with learning, memory and aging but also in autoimmune diseases, including rheumatoid arthritis (RA).

### NGF AND THE PERIPHERAL NERVOUS SYSTEM

• NGF functions as a retrograde trophic messenger between target tissues and their innervating nerve cells. NGF and NGF mRNA have been detected in peripheral tissues of various mammalian species, whereas NGF receptor mRNA is mostly produced by the corresponding sensory and sympathetic ganglia (2,15,16). NGF exerts its neurotrophic activity in re-

sponse to ligand-receptor binding followed by internalization and retrograde transport of the NGF-receptor complex to the neuronal cell body. Recent studies showed the presence of two different types of NGF receptors: p75 acting as a low-affinity receptor ( $K_d=10^9M$ ) and p75<sup>NTR</sup> high-affinity NGF receptor ( $K_d=10^{-11}M$ ) (17-19).

Daily injection of purified NGF into neonatal rodents results in an increase in volume of sympathetic ganglia, hypertrophy of sensory ganglia, and in an augmentation of their neurotransmitters within NGF target cells (2,15,20-22). The effect of NGF in sympathetic nerve cells is even more dramatically documented by the finding that removal of circulating NGF in living animals, via administration of specific NGF antibodies, results in death of sympathetic neurons, leading to what has been known for years as immunosympathectomy (23).

### NGF AND THE CENTRAL NERVOUS SYSTEM

- The actions of NGF are not limited to the peripheral nervous system. Numerous studies published in the last ten years have shown that NGF is produced in the central nervous system (5,6), that cholinergic basal forebrain neurons (CBFN) bear NGF receptors (17,19,24,25), and that these neurons are highly receptive to the action of NGF. NGF and its receptor are axonally transported retrogradely from cortex and hippocampus to CBFN, where they exert a trophic action. It has also been clearly demonstrated that the degeneration of CBFN following transection of the septo-hippocampal pathway is markedly reduced by exogenous administration of NGF. These studies support the notion that NGF exerts a functional effect on these brain neurons (26) not only during development but also in adult and aged life. Moreover, the widespread presence of NGF mRNA throughout cells of the central nervous system suggests other functions of this protein within the brain (13,14,27-30).

### NGF AND THE ENDOCRINE SYSTEM

- Apart from its well-known roles in the survival and development of peripheral sympathetic and sensory neurons and in CBFN, NGF is also known to take part in the regulation of specific neuroendocrine functions (13,30-34). In fact, it has been shown that both NGF and its mRNA increase in the hypothalamus following stressful events (11-14). Moreover, the injection of NGF antibodies into rat fetuses results in a marked neuroendocrine deficit in postnatal life, including atrophied peripheral sympathetic and sensory ganglia (31). Deleterious effects of NGF deprivation during fetal life have also been reported in rats and guinea pigs (32,33). Furthermore, maternal exposure to NGF antibodies causes, in newborn pups, loss of body weight, sensory deficits, and high mortality, probably associated to neuroendocrine and immune

alterations (30). In this context, it is worth mentioning that exogenous NGF administration acts on the hypothalamic-pituitary-adrenal axis by influencing, most probably, the release of hypothalamic hormones (35).

### HGF AND THE IMMUNE SYSTEM

- There is ample evidence that NGF is able to exert a wide variety of effects on cells of the immune system (36-47). These studies demonstrated that *in vivo* administration of NGF to neonatal rats causes a widespread increase in the size and number of MC in several peripheral tissues (36). Exposure of spleen cells to NGF antibodies results in a cytological alteration and a reduction of MC numbers. Recently, it has been shown that MC bear NGF receptors and that NGF induces degranulation and histamine release from isolated peritoneal MC (48-51). That NGF plays a role in the immune system is also suggested by findings which show that administration of NGF in young rats prior to and after immunization with sheep erythrocytes results in an enhancement of T-lymphocyte-dependent antibody synthesis (52).

NGF has also been shown to be involved in inflammatory responses. The observation that inflammation causes an increase in NGF levels was first reported by Levi-Montalcini and Angeletti, who showed that carrageenan-induced granuloma causes a dramatic local increase in NGF levels, far above that found with sarcoma 180 (53). These studies were recently confirmed and extended, demonstrating that cells normally present in inflammatory tissues, such as MC and lymphocytes, express NGF receptors (46,50,54) and are receptive to the action of NGF (36,37). These findings along with the observation that NGF accumulates at the site of the lesion and inflammation and that cytokines, such as interleukin-1 $\beta$  (IL-1 $\beta$ ) and tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), are potent inducers of NGF secretion (55-57) suggest a functional link between cytokines and NGF in certain inflammatory diseases.

### HGF AND AUTOIMMUNE DISEASES

- The observation that the concentration of NGF is enhanced in inflammatory response, along with the emerging evidence that cells of immune system lineage are able to respond to and/or synthesize NGF (30) led to the hypothesis that this molecule might be involved in autoimmune diseases. Thus, it was assumed that autoimmune diseases characterized by abnormal activation of the immune system and by alteration of numerous biologically active mediators may influence the synthesis and release of NGF. Indeed, recent studies carried out in our laboratory demonstrated elevated NGF levels in a large number of autoimmune diseases (58-66). For example, NGF has been found in the synovial fluid of RA patients and other forms of chronic arthritis, as well as in the

synovium of animals with pharmacologically-induced arthritis (58-60). In the latter instance, destruction of peripheral sympathetic innervation reduced both the inflammation and the increased level of NGF caused by carrageenan injection (59). More recently, it was demonstrated that the synovium of transgenic arthritic mice carrying and expressing the human TNF- $\alpha$  gene accumulates elevated levels of NGF. Interestingly, subcutaneous injection of NGF antibodies in these transgenic animals attenuated the wasting effect of TNF- $\alpha$  (61).

Basal levels of NGF were found to be altered in the presence of other autoimmune diseases. For example, the concentration of NGF increases in the cerebrospinal fluid of patients with multiple sclerosis (MS) (62) and in the brain of rats affected by experimental allergic encephalomyelitis (EAE), an animal model of MS (63). A significant increase in NGF levels was observed in the sera of patients with systemic lupus erythematosus (SLE) and in dermis of patients affected by systemic sclerosis (SSc) (64,65). In most of these diseases, the increase of NGF was invariably associated with the accumulation of MC. In view of the fact that MC secrete NGF (67), express NGF mRNA and bear low- and high-affinity NGF receptors (50,54), and since both NGF and MC are up-regulated during inflammation, it is highly possible that either singly or cooperatively they may be functionally involved in these diseases (68-75).

#### MGF AND MAST CELLS

• Under normal conditions MC are found in diverse tissues and organs, including nervous tissue, skin, gastrointestinal and respiratory tracts, vascular wall and endocrine glands (68-70,76). MC have been identified in a variety of inflammatory states, such as SLE, mixed connective tissue disease, SSc, atopic dermatitis, MS, and EAE. They respond to and produce a variety of cytokines, such as IL-1,3,4,5,6, TNF- $\alpha$ , interferon- $\gamma$ , granulocyte-macrophage colony stimulating factor and TNF- $\beta$  (69,71). In addition, peptides derived from granulocytes, platelets, and mononuclear cells, as well as complement-derived peptides and certain proteases are able to stimulate MC secretion. MC can be activated by nervous and immunological mediators such as neuropeptides (e.g. substance P) or immunoglobulin E (IgE) and are known to be involved in the development of immediate and delayed hypersensitivity reactions (75,77-79).

The observation that MC respond to NGF by increasing in numbers and by releasing biological mediators (30,48,67) and that they accumulate in the synovium of patients affected by arthritis and other inflammatory disorders (72-75), suggested that attention be focused on the role of NGF in autoimmune diseases. As a first approach to carry out this studies, we selected human and rodent arthritis. We therefore decided to

study the relationship of MC and NGF in rodents and humans affected by this type of disease. Our data clearly indicate that human and experimentally-induced arthritis is characterized by an increase in MC numbers and that MC distribution is correlated to NGF levels (65,66). Since MC produce cytokines (69,71) and cytokines are known to influence the synthesis of NGF (55-57), the results of time-course studies will help to elucidate the functional relationship between NGF, MC and cytokines in autoimmune diseases.

#### MGF, MAST CELLS AND ARTHRITIS

• RA is a multisystem, chronic, inflammatory disease of not well-defined origin (72-75). The inflammatory conditions are characterized by synovial hypertrophy and hyperplasia, and mixed cell infiltration, including numerous MC (78-81). The observation that these cells are associated with development of an inflammatory infiltrate in arthritis suggested that the identification of the mechanism leading to the accumulation of MC might contribute to the development of better therapeutic strategies for arthritis.

The fact that MC are multifunctional cells which store and release numerous biological mediators, including histamine, heparin, cytokines (71) and NGF (67,81), raised the important question of what role NGF plays during the course of inflammatory diseases. Our previous studies showing that injection of NGF into neonatal rats causes an increase of MC led to the hypothesis that also in arthritis, the presence of MC is the result of an overproduction of NGF. During synovial inflammation the increase of NGF precedes the accumulation of MC. However, recent observations that injection of highly purified NGF into the knee joint does not induce inflammation and that MC are able to release NGF (66,67) question this hypothesis and raise the possibility that MC may contribute to up-regulation of synovial NGF. Based on the present findings, one possible explanation is that the presence of high NGF levels at the site of inflammation may serve as a regulatory mechanism for a subpopulation of immunocompetent cells that are necessary for healing processes. The fact that MC can alter lymphocyte function favoring suppression of cytotoxic effects, whilst recruiting phagocytic leukocytes, supports this hypothesis (74). However, this hypothesis is consistent with the observations that IL-4 released by both MC/basophils and T-lymphocytes inhibits the production of proinflammatory cytokines such as IL-1, IL-6, IL-8 and TNF- $\alpha$  (77). It is therefore also possible that NGF acts through the action of IL-4 which is also a potent inhibitor of synthesis by monocytes of both prostaglandin-E<sub>2</sub> and collagenase. Indeed, IL-4 was able to reduce disease activities and progression in various arthritic models (77).

Elevated concentrations of NGF have also been observed after

a number of degenerative events within and outside the nervous system and have been associated to neurite regeneration mechanisms (82-85). It is therefore possible that similarly to the nervous system, the increase of NGF and/or MC occurring in certain tissues during autoimmune diseases is linked to protective and reparative mechanisms (86-88). The observation that the level of NGF in SSc, MS and in EAE is high during the acute phase and low during the remission one (62,63,66) is consistent with this hypothesis. Since NGF is known to act on cells of the nervous, immune and endocrine systems (30,76,85), it is possible that during inflammation its role is to participate in peripheral tissue repair and to reestablish physiological conditions.

In this context is also worth mentioning that MC degranulation through release of heparin may also have antiinflammatory effects. For example, inhibition of T-lymphocyte heparanase by heparin has been shown to prevent T-cell migration, whereas treatment with low doses of heparin has been found to inhibit adoptively transferred encephalomyelitis (89,90). Heparin has also been demonstrated to modulate cell proliferation, wound healing, neurite outgrowth, and inflammation (91). Moreover, the potential role of peripheral MC may be related to connective tissue formation in development or repair and in defence against parasite infection (84,85).

## CONCLUSIONS

- Several autoimmune diseases, including arthritis, are associated with the release of numerous biologically active mediators from MC and since MC mediators are known to exert not only deleterious (69,72-74), but also beneficial effects (84-93), a better understanding of the functional correlation between NGF and MC might be of great value for the development of therapeutic strategies for treating joint inflammatory diseases.

## ACKNOWLEDGEMENTS

- This study was supported by Target Project FATMA, Subproject Stress, and Target Project on Aging from the Consiglio Nazionale delle Ricerche (CNR).

## REFERENCES

1. Levi-Montalcini R, Angeletti PU. The nerve growth factor. *Physiol Rev* 1968; 48: 534-569
2. Levi-Montalcini R. The nerve growth factor 35 years later. *Science* 1987; 237: 1154-1162
3. Barka T. Biologically active polypeptides in submandibular glands. *J Histochem Cytochem* 1980; 28: 836-859
4. Harper GP, Glanville RV, Thoenen H. The purification of nerve growth factor from bovine seminal plasma. *J Biol Chem* 1982; 257: 8541
5. Gnahn H, Hefty F, Heumann R, Schwab ME, Thoenen H. NGF-mediated increase of choline acetyltransferase (ChAT) in the neonatal rat forebrain, evidence for a physiological role of [3-NGF in brain? *Dev Brain Res* 1983; 3: 229-238
6. Thoenen H, Bandtlow C, Heumann R. The physiological function of nerve growth factor in the central nervous system: comparison with the periphery. *Physiol Biochem Pharmacol* 1987; 109: 146-171
7. Bocchini G, Angeletti PU. The nerve growth factor: purification as a 30,000-molecular weight protein. *Proc Natl Acad Sci USA* 1969; 64: 787-794
8. Goedert M, Fine A, Hunt SP, Ullrich. A nerve growth factor mRNA in peripheral and central rat tissues and in the human central nervous system: Lesion effect in the rat brain and levels in Alzheimer's disease. *Mol Brain* 1986; 1: 85-92
9. Green LA, Shooter EM. The nerve growth factor: biochemistry, synthesis and mechanism of action. *Ann Rev Neurosci* 1980; 3: 353-402
10. Snider WD, Johnson EM. Neurotrophic molecules. *Ann Neurol* 1989; 26: 489-206
11. Aloe L, Alleva E, Bohm A, Levi-Montalcini R. Aggressive behaviour induces release of nerve growth factor from mouse salivary gland into the bloodstream. *Proc Natl Acad Sci USA* 1986; 83: 6184-6187
12. Alleva E, Aloe L. Physiological roles of NGF in adult rodents: a biobehavioral perspective. *Int J Comp Psychol* 1989; 2: 147-163
13. Aloe L, Alleva E, De Simone R. Changes of NGF level in mouse hypothalamus following intermale aggressive behavior: biological and immunohistochemical evidence. *Behav Brain Res* 1990; 39: 53-61
14. Spillantini MG, Aloe L, Alleva E, De Simone R, Goedert M, Levi-Montalcini R. Nerve growth factor mRNA and protein increase in hypothalamus in a mouse model of aggression. *Proc Natl Acad Sci USA* 1989; 86: 8555-8559
15. Goedert M, Stoeckel K, Otten U. Biological importance of the retrograde transport of nerve growth factor in sensory neurons. *Proc Natl Acad Sci USA* 1981; 78: 5895-5898

16. Shelton DL, Reichard LF. Expression of nerve growth factor gene correlates with the density of sympathetic innervation of effector organs. *Proc Natl Acad Sci USA* 1984; 81: 7951-7955
17. Meakin SO, Shooter EM. The nerve growth factor family of receptors. *Trends Neurosci* 1992; 15:323-331
18. Barde YA. Trophic factors and neuronal survival. *Neuron* 1989; 2: 1525-1534
19. Barbacid M. Nerve growth factor: a tale of two receptors. *Oncogene* 1993; 8: 2033-2042
20. Lindsay RM, Harmar AJ. Nerve growth factor regulates expression of neuropeptide genes in adult sensory neurons. *Nature* 1989; 337: 362-364
21. Donnerer J, Schuligoi R, Stein C. Increased content and transport of substance P and calcitonin gene-related peptide in sensory nerves innervating inflamed tissue: evidence for a regulatory function of nerve growth factor *in vivo*. *Neuroscience* 1992; 49: 693-698
22. Lewin GR, Ritter AM, Mendell LM. Nerve growth factor-induced hyperalgesia in the neonatal and adult rat. *J Neurosci* 1993; 13: 2136-2148
23. Levi-Montalcini R, Angeletti PU. Immunosympathectomy. *Pharmacol Rev* 1966; 18: 619-628
24. Shelton DL, Reichard LF. Studies in the expression of nerve growth factor gene in the central nervous system: level and regional distribution of NGF mRNA suggest that NGF functions as a trophic factor for several distinct populations of neurons. *Proc Natl Acad Sci USA* 1986; 83: 2714-2718
25. Raivich G, Kreitzberg GW. The localization and distribution of high affinity beta nerve growth factor binding sites in the central nervous system of rats. A light microscopic autoradiographic study using <sup>25</sup>I-NGF. *Neuroscience* 1978; 20: 923-934
26. Hefty F. Nerve growth factor promotes survival of septal cholinergic neurons after fimbrial transections. *J Neurosci* 1986; 6: 2155-2162
27. Lara HE, McDonald JK, Ojeda SR. Involvement of nerve growth factor in female sexual development. *Endocrinology* 1990; 126: 364-375
28. Ojeda SR, Hill DF, Katz KH. The genes encoding nerve growth factor and its receptor are expressed in the developing female rat hypothalamus. *Mol Brain Res* 1991; 9: 47-55
29. Luppi P, Levi-Montalcini R, Bracci-Laudiero L, Bertolini A, Arletti R, Tavernari D, *et al*. NGF is released into plasma during human pregnancy: an oxytocin-mediated response? *NeuroReport* 1993; 4: 1063-1065
30. Levi-Montalcini R, Aloe L, Alleva E. A role for nerve growth factor in nervous, endocrine and immune systems. *Prog Neuroendocrine Immunol* 1990; 3: 1-10
31. Aloe L, Cozzari C, Calissano P, Levi-Montalcini R. Somatic and behavioral postnatal effects of fetal injections of nerve growth factor antibodies in the rat. *Nature* 1981; 291:413-415
32. Johnson EM Jr, Osborne PA, Rydel RE, Schmidt RE, Pearson J. Characterization of the effects of autoimmune nerve growth factor deprivation in the developing guinea pig. *Neuroscience* 1993; 8: 631-642
33. Yip HK, Rich KM, Lampe PA, Johnson EM Jr. The effects of nerve growth factor and its antiserum on the postnatal development and survival after injury of sensory neurons in rat dorsal root ganglia. *J Neurosci* 1984; 4: 2986-2992
34. Missale C, Boroni F, Losa M, Giovanelli M, Zanellato A, Dal Toso R, *et al*. Nerve growth factor suppresses the transforming phenotype of human prolactinomas. *Proc Natl Acad Sci USA* 1993; 90: 7961-7965
35. Otten U, Baumann JB, Girard J. Stimulation of the pituitary-adrenocortical axis by nerve growth factor. *Nature* 1979; 282: 413-414
36. Aloe L, Levi-Montalcini R. Mast cells increase in tissues of neonatal rats injected with the nerve growth factor. *Brain Res* 1977; 133: 358-366
37. Thorpe LW, Perez-Polo JR. The influence of nerve growth factor on the *in vitro* proliferative response of rat spleen lymphocytes. *J Neurosci Res* 1987; 18: 134-139
38. Otten U, Ehrhard P, Peck R. Nerve growth factor induces growth and differentiation of human B lymphocytes. *Proc Natl Acad Sci USA* 1989; 86: 10059-10063
39. Brodie C, Gelfand EW. Functional nerve growth factor receptors on human B lymphocytes. Interaction with IL-2. *J Immunol* 1992; 148: 3492-3497
40. Kimata H, Yoshida A, Ishioka C, Kusunoki T, Hosoi S,

- Mikawa H. Nerve growth factor specifically induces human IgG4 production. *Eur J Immunol* 1991; 21: 137-141
41. Thorpe LW, Werrbach-Perez K, Perez-Polo JR. Effects of nerve growth factor expression on interleukin-2 receptors on cultured human lymphocytes. *Ann NY Acad Sci* 1987; 496: 310-311
  42. Matsuda H, Denurg JA. Nerve growth factor promotes human hemopoietic colony growth and differentiation. *Proc Natl Acad Sci USA* 1988; 85: 6508-6512
  43. Gee AP, Boyle MDP, Munger KL, Lawman MJP, Young M. Nerve growth factor: stimulation of polymorphonuclear leukocyte chemotaxis *in vitro*. *Proc Natl Acad Sci USA* 1983; 80: 7215-7218
  44. Boyle MDP, Lawman MJP, Gee AP, Young M. Nerve growth factor: a chemotactic factor for polymorphonuclear leukocytes *in vivo*. *J Immunol* 1985; 134: 564-568
  45. Morgan B, Thorpe LW, Marchetti D, Perez-Polo JR. Expression of nerve growth factor receptors by human peripheral blood mononuclear cells. *J Neurosci Res* 1989; 23:41-45
  46. Ehrhard PB, Ganter U, Bauer J, Otten U. Expression of functional trk protooncogene in human monocytes. *Proc Natl Acad Sci USA* 1993; 90: 5423-5427
  47. Ehrhard PB, Erb P, Graumann U, Otten U. Expression of nerve growth factor and nerve growth factor receptor tyrosine kinase Trk in activated CD4-positive T-cell clones. *Proc Natl Acad Sci USA* 1980;90: 10984-10988
  48. Bruni A, Bigon E, Borato E, Mietto L, Leon A, Toffano G. Interaction between nerve growth factor and lysophosphatidylserine on rat peritoneal mast cells. *FEES Lett* 1982; 138: 190-192
  49. Pearce FL, Thompson HL. Some characteristics of histamine secretion from rat peritoneal mast cells stimulated with nerve growth factor. *J Physiol* 1986; 372: 379-393
  50. Aloe L. The effect of nerve growth factor and its antibody on mast cells *in vivo*. *J Neuroimmunol* 1988; 18: 1-12
  51. Aloe L, De Simone R. NGF primed spleen cells injected in brain of developing rats differentiate into mast cells. *M J Dev Neurosci* 1989; 7: 565-573
  52. Manning PT, Russell JH, Simmons B, Johnson EM Jr. Protection from guanethidine-induced neuronal destruction by nerve growth factor: effects of NGF on immune function. *Brain Res* 1985; 340: 61-69
  53. Levi-Montalcini R, Angeletti PU. Biological properties of a nerve growth factor promoting protein and its antiserum. In: Kety Elkees SS, editor. Fourth Internat. Neurochem. Symposium. New York, Pergamon Press, 1968; 362-367
  54. Horigome K, Pryor JC, Bullock ED, Johnson EM Jr. Mediator release from mast cells by nerve growth factor. Neurotrophin specificity and receptor mediation. *J Biol Chem* 1993; 268: 14881-14887
  55. Gasser UE, Weskamp G, Otten U, Dravid AR. Time course of the elevation of nerve growth factor (NGF) content in the hippocampus and septum following lesions of the septohippocampal pathway in rats. *Brain Res* 1986; 376:351-356
  56. Friedman WJ, Larkfors L, Ayer-LeLievre C, Ebendal T, Olson L, Persson H. Regulation of p-nerve growth factor expression by inflammatory mediators in hippocampal cultures. *J Neurosci Res* 1990; 27: 374-382
  57. Hattori A, Tanaka E, Murase K, Ishida N, Chatani Y, Tsujimoto H, *et al*. Tumor necrosis factor stimulates the synthesis and secretion of biologically active nerve growth factor in non-neuronal cells. *J Biol Chem* 1993; 268: 2577-2582
  58. Aloe L, Tuveri MA, Carcassi U, Levi-Montalcini R. Nerve growth factor in the synovial fluid of patients with chronic arthritis. *Arthr Rheum* 1992; 35: 351-355
  59. Aloe L, Tuveri MA, Levi-Montalcini R. Studies on carageenan-induced arthritis in adult rats: presence of nerve growth factor and role of sympathetic innervation. *Rheumatol Int* 1992; 12: 213-216
  60. Aloe L, Tuveri MA, Levi-Montalcini R. Nerve growth factor and distribution of mast cells in the synovium of adult rats. *Clin Exp Rheumatol* 1992; 10: 203-204
  61. Aloe L, Probert L, Kollias G, Bracci-Laudiero L, Spillantini MG, Levi-Montalcini R. The synovium of transgenic arthritic mice expressing human tumor necrosis factor contains a high level of nerve growth factor. *Growth Factors*. In press
  62. Bracci-Laudiero L, Aloe L, Levi-Montalcini R, Buttinelli C, Schilter D, Gillessen S, *et al*. Multiple sclerosis patients express increased levels of [3-nerve growth factor in cerebrospinal fluid. *Neurosci Lett* 1992; 147: 9-12
  63. Micera A, De Simone R, Aloe L. Elevated levels of nerve growth factor in the thalamus and spinal cord of rats af

- fectured by experimental allergic encephalomyelitis. *Arch Ital Biol*. In press
64. Bracci-Louder L, Aloe L, Levi-Montalcini R, Galeazzi M, Schilter D, Scully JL, *et al*. Increased levels of NGF in sera of systemic lupus erythematosus patients. *Neuro Report* 1993; 4: 563-565
  65. Tuveri MA, Passiu G, Mathieu A, Aloe L. Nerve growth factor and mast cell distribution in the skin of patients with systemic sclerosis. *Clin Exp Rheumatol* 1993; 11: 319-322
  66. Aloe L, Probert L, Kollias G, Micera A, Tirassa P. Effect of NGF antibodies on mast cell distribution, histamine and substance P levels in the knee joint of TNF-arthritis transgenic mice. *Rheumatol Int* 1995; 14: 249-252
  67. Leon A, Buriani A, Dal Toso R, Fabris M, Romanello S, Aloe L. Mast cells synthesize, store, and release nerve growth factor. *Proc Natl Acad Sci USA* 1994; 91: 3739-3743
  68. Bienstock J, Tomioka M, Matsuda H, Stead RH, Quinonez, Simon GT, *etal*. The role of mast cells in inflammatory processes: evidence for nerve/mast cell interactions. *Int Arch Allergy Appl Immunol* 1987; 82: 238-243
  69. Galli SJ. New concepts about the mast cell. *New Engl J Med* 1993; 328:257-265
  70. Johnson D, Krenger W. Interactions of mast cells with the nervous system - recent advances. *Neurochem Res* 1992; 9: 939-951
  71. Gordon JR, Burd PR, Galli SJ. Mast cells as a source of multifunctional cytokines. *Immunol Today* 1990; 11:458-464
  72. Malone DG, Metcalfe DD. Mast cells and arthritis. *Ann Allergy* 1988; 61: 27-30
  73. Mican JM, Metcalfe DD. Arthritis and mast cell activation. *J Allergy Appl Immunol* 1990; 86: 677-683
  74. Wasserman SI. The mast cell and synovial inflammation. Or, what's a nice cell like you doing in a joint like this,?! *Arthritis Rheum* 1984; 27: 841-844
  75. Gruber BL. Immunoglobulin E, mast cells, endogenous antigens, and arthritis. *Rheum Dis Clin North America* 1991; 17: 333-342
  76. Chaldakov GN, Ghenev PI, Andonov M, Valchanov K, Tonchev A, Pancheva R. Neural-immune-effector (NIE) cross-talk in vascular trophobiology: proposal for new and not yet exploited purinergic regulatory mechanisms. *Biomed Rev* 1994; 3: 81-86
  77. Miossec P. Interleukin 4. A potential anti-inflammatory agent. *Rev Rheum* 1993; 60: 87-91
  78. Burd PR, Rogers HW, Gordon JR, *et al*. Interleukin-3 dependent and independent mast cells stimulated with IgE and antigen express multiple cytokines. *J Exp Med* 1989; 170: 245-254
  79. Plaut M, Pierce JH, Watson CJ, *et al*. Mast cell lines produce lymphokines in response to cross-linkage of Fc epsilon RI or to calcium ionophores. *Nature* 1989; 339: 64-67
  80. Tsai M, Gordon JR, Galli SJ. Mast cells constitutively express transforming growth factor-(3 mRNA [abstract]. *FASEB J* 1990; 4: 1944A
  81. Aloe L, Skaper SD, Leon A, Levi-Montalcini R. Nerve growth factor and autoimmune diseases. *Autoimmunity* 1994; 19: 141-150
  82. Gall CM, Isackson PJ. Limbic seizures increase neuronal production of mRNA for nerve growth factor. *Science* 1994; 242: 758-761
  83. Kromer LF. Nerve growth factor treatment after brain injury prevent neuronal death. *Science* 1987; 235: 214-216
  84. Stead RH, Tomioka M, Quinonez G, Simon GT, Felten SY, Bienstock J. Intestinal mucosal mast cells in normal and nematode-infected intestines are in intimate contact with peptidergic nerves. *Proc Natl Acad Sci USA* 1987; 84: 2975-2979
  85. McKay DM, Bienstock J. The interaction between mast cells and nerves in the gastrointestinal tract. *Immunol Today* 1994; 15: 533-538
  86. Bischoff SC, Dahinden CA. Effect of nerve growth factor on the release of inflammatory mediators by mature human basophils. *Blood* 1992; 79: 2662-2669
  87. Heumann R, Korsching S, Bandtlow C, Thoenen H. Changes of nerve growth factor synthesis in non-neuronal cells in response to sciatic nerve transection. *J Cell Biol* 1987; 104: 1623-1631
  88. Lindholm D, Heumann R, Meyer M, Thoenen H. Interleukin-1 regulates synthesis of nerve growth factor

- in non-neuronal cells of the rat sciatic nerve. *Nature* 1987; 330:658-659
89. Lider O, Baharav E, Mekori YA, Miller T, Naparstek Y, Vlodaysky I, Cohen IR. Suppression of experimental autoimmune diseases and prolongation of allograft survival by treatment of animals with low doses of heparin. *J Clin Invest* 1989; 752-756
90. Lider O, Mekori YA, Miller T, Bar-Tana R, Vlodaysky I, Baharav E, *et al.* Inhibition of T-lymphocyte heparinase by heparin prevents T-cell migration and T-cell-mediated immunity. *Eur J Immunol* 1990 20: 493-499
91. Tyrrell DJ, Kilfeather S, Page CP. Therapeutic uses of heparin beyond its traditional role as an anticoagulant. *Trends Pharmacol Sci* 1995; 16: 198-204
92. Franzen N, Grassefifar R, Malcherek P. Experimental mast cell activation improves connective tissue repair in perforated rat mesentery. *Agents Actions* 1990; 33: 371-377
93. Grant JA, Alam R, Lett-Brown MA. Histamine-releasing factor and inhibitors: historical perspective and possible implications in human illness. *J Allergy Clin Immunol* 1991; 88: 683-93

*Received 28 July 1995*

*Accepted 29 August 1995*

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