

EFFECTS OF ARONIA MELANOCARPA FRUIT JUICE ON LIPID AND BONE METABOLISM IN OVARIECTOMIZED RATS

Antoaneta Georgieva¹, Milena Todorova¹, Miroslav Eftimov¹, Krasimir Kuzmanov²,
Vasilena Kuzmanova³, Atanas Kuzmanov³, Mila Vlaskovska⁴, Stefka Valcheva-Kuzmanova¹

¹Department of Pharmacology and Clinical Pharmacology and Therapeutics,
Faculty of Medicine, Medical University of Varna

²Vivarium, Medical University of Varna

³Student, Faculty of Dental Medicine, Medical University of Varna

⁴Department of Pharmacology and Toxicology, Faculty of Medicine,
Medical University of Sofia

ABSTRACT

INTRODUCTION: The ovariectomized (OVX) rat is a gold-standard model to mimic the changes in the female organism during menopause. *Aronia melanocarpa* fruits are very rich in polyphenols and exert many beneficial effects in animal models and clinical trials.

AIM: The aim of the present study was to investigate the effects of *Aronia melanocarpa* fruit juice (AMFJ) on lipid metabolism and bone mineral density (BMD) in OVX rats.

MATERIALS AND METHODS: Four groups of female Wistar rats were formed, each consisting of 14 animals—sham operated (SO), OVX (treated with distilled water), as well as OVX+AMFJ₅, and OVX+AMFJ₁₀ (OVX treated with 5 and 10 mL/kg AMFJ, respectively). The animal treatment began 2 weeks after the operation. After a three-month treatment period, body weight (BW), total fat (TF) and retroperitoneal fat (RPF), as well as serum total cholesterol were measured. The indices TF/BW and RPF/BW were calculated. Femur BMD was determined as well. The statistical analysis used was one-way ANOVA.

RESULTS: Compared to SO animals, OVX rats showed a statistically significant increase in TF ($p < 0.05$), RPF ($p < 0.01$), TF/BW ($p < 0.05$), and RPF/BW ($p < 0.01$). Plasma cholesterol levels of OVX animals were increased and their BMD was decreased if compared to SO rats, but these changes were not statistically significant. Compared to OVX rats, AMFJ did not affect lipid accumulation and cholesterol levels but at the dose of 10 mL/kg significantly increased BMD ($p < 0.05$ vs. OVX).

CONCLUSION: *Aronia melanocarpa* fruit juice was not able to prevent the fat accumulation in OVX rats, but managed to restore their BMD.

Keywords: *Aronia melanocarpa*, polyphenols, lipid metabolism, bone mineral density, ovariectomy, rats

Address for correspondence:

Antoaneta Georgieva
Faculty of Medicine
Medical University of Varna
55 Marin Drinov St
9002 Varna

e-mail: antoaneta.georgieva@mu-varna.bg

Received: February 2, 2022

Accepted: February 16, 2022

INTRODUCTION

Menopause is a physiological condition resulting from the decline in ovarian estrogen production. This hormonal deficit has some serious consequences for female health. Postmenopausal estrogen deficit often results in dyslipidemia (decreased HDL cholesterol, as well as increased total cholesterol and LDL cholesterol) and increased bone turnover with osteoporosis and increased fracture risk (1). Menopause also increases the risk of obesity and often causes abdominal fat accumulation (2). The ovariectomized (OVX) rat is a gold-standard animal model that mimics the clinical findings in menopause (3).

Table 1. Aronia melanocarpa fruit juice (AMFJ) ingredients and methods of their determination; GAE—gallic acid equivalent, HPLC—high-performance liquid chromatography; the content of the juice was determined by the authors (6)

Ingredient	Content	Method of Determination
Total phenols	5461 GAE/L	Folin-Ciocalteu procedure (13)
Total proanthocyanidins	3122.5 mg/L	Gravimetrically according to the procedure described by Howell et al. (14)
Cyanidin 3-galactoside	143.7 mg/L	HPLC
Cyanidin 3-arabinoside	61.7 mg/L	HPLC
Cyanidin 3-glucoside	4.4 mg/L	HPLC
Cyanidin 3-xyloside	11.6 mg/L	HPLC
Chlorogenic acid	585 mg/L	HPLC
Neochlorogenic acid	830 mg/L	HPLC

Aronia melanocarpa (Michx) Elliot, called also black chokeberry, belongs to the *Rosaceae* plant family. Its fruits are extremely rich in polyphenols, especially proanthocyanidins, flavonoids (anthocyanins, quercetin glycosides) and phenolic acids (chlorogenic and neochlorogenic) (4–6).

Aronia melanocarpa hot water extracts were shown to improve the lipid profile and bone mineral density (BMD) in OVX rats (7). Other polyphenols and polyphenol-rich plant products such as blueberry powder (8), grape proanthocyanidins (9), chlorogenic acid (10,11), and quercetin (12) were able to increase BMD in rats with different osteoporosis models (induced by OVX, glucocorticoids or retinoic acid).

AIM

The aim of the present study was to investigate the effect of *Aronia melanocarpa* fruit juice (AMFJ)

on fat accumulation, plasma cholesterol levels, and femur BMD in rats with OVX-induced estrogen deficit.

MATERIALS AND METHODS

Aronia Melanocarpa Fruit Juice

Aronia melanocarpa Elliot fruits were grown in the Balkan Mountains in Bulgaria. To prepare AMFJ, they were crushed and squeezed. Then, the juice was filtered, preserved with potassium sorbate (1.0 g/L), and stored at 0°C until the experiment (6). The contents of phenolic substances in AMFJ and the method of their determination are given in Table 1.

Animals and Operation

Female rats were allocated into four groups consisting of 14 animals each—sham-operated (SO) and three groups of OVX rats: OVX, OVX+AMFJ₅, and OVX+AMFJ₁₀—receiving 5 and 10 mL/kg AMFJ, respectively. Rats were four months old at the beginning of the experiment and sexually naïve. On the day of the operation, anesthesia with ketamine 30 mg/kg and xylazine 30 mg/kg was performed and animals were fixed. The abdominal hair was removed and the abdomen was disinfected with iodine. After that a midline incision was performed. The abdominal cavity was sewed back immediately in SO rats. In OVX rats, the ovaries were isolated with consecutive clamping of uterine tubes and tying a thread around the oviduct and its blood vessels. After this, the abdominal wall was closed, and prophylaxis with ceftazolin 200 mg/kg i.p. was performed. There was a recovery period of two weeks after the operation. Af-

ter that, the daily oral treatment began. The experimental substances were applied to rats, using an orogastric tube. Distilled water (10 mL/kg) was given to SO and OVX rats. OVX+AMFJ₅ and OVX+AMFJ₁₀ animals were treated respectively with AMFJ 5 mL/kg (diluted with water to a volume of 10 mL/kg) and AMFJ 10 mL/kg. Animal care and all experiments were in conformity with national laws and policies as well as with the international guidelines (EU Directive, 2010/63/EU for animal experiments).

Three months after the operation, several behavioral tests were performed. On the last day, animal body weight (BW) was measured. Then, after diethyl ether anesthesia rats were sacrificed and blood was collected from the sublingual veins to prepare serum for biochemical investigations. Then fat deposits (retroperitoneal, mesenteric and perigonadal) were dissected and measured as total fat (TF). Retroperitoneal fat (RPF) was separately weighed. The indices TF/BW and RPF/BW were calculated. Fe-

considered significant. GraphPad Prism 5 statistical software was used.

RESULTS

Lipid Metabolism

Data regarding lipid metabolism are presented in Table 2. All OVX animals (both treated with distilled water and AMFJ) had significantly higher fat deposits if compared to SO rats ($F_{3,50}=3.92$, $p<0.05$ for TF; $F_{3,49}=5.39$, $p<0.01$ for RPF; $F_{3,51}=3.50$, $p<0.05$ for TF/BW index and $F_{3,49}=5.78$, $p<0.01$ for RPF/BW index). *Aronia melanocarpa* fruit juice-treated groups had fat deposits which were not significantly different than those of OVX animals.

The mean cholesterol level of OVX group was higher than that of SO animals with no statistical significance. Compared to OVX group, the cholesterol levels of AMFJ-treated groups were also not significantly different.

Table 2. Total (TF, g), retroperitoneal fat (RPF, g), TF/BW and RPF/BW indices, total plasma cholesterol (TC, mmol/L); * $p < 0.05$, ** $p < 0.01$ vs. SO; BW—body weight

	TF (g)	RPF (g)	TF/BW	RPF/BW	TC (mmol/L)
SO	9.4 ± 1.2	1.6 ± 0.2	0.04 ± 0.005	0.006 ± 0.001	2.1 ± 0.1
OVX	15.4 ± 1.8*	3.3 ± 0.4**	0.05 ± 0.005*	0.012 ± 0.001**	2.4 ± 0.1
OVX + AMFJ ₅	14.2 ± 1.5*	3.1 ± 0.4**	0.05 ± 0.005	0.011 ± 0.001**	2.3 ± 0.1
OVX + AMFJ ₁₀	15.0 ± 1.3*	3.2 ± 0.3**	0.05 ± 0.004*	0.012 ± 0.001**	2.3 ± 0.1

murs were removed, cleared from all soft tissues and frozen.

Biochemical Measurements

Serum levels of total cholesterol were measured spectrophotometrically (spectrophotometer CE2021, Cecil Instruments Ltd, UK). The analysis was performed using kits from Biomaxima (Poland) according to manufacturer's instructions.

X-Ray Absorptiometry (DXA)

Femur BMD was measured by dual energy X-ray absorptiometry (DXA) using a computer program for small subjects.

Statistical Analyses

Statistical analyses were performed by one-way analysis of variance (one-way ANOVA) with post-hoc Dunnett's multiple comparison test. Data were presented as mean ±SEM. A value of $p<0.05$ was

Bone Mineral Density

The mean BMD of rats belonging to SO, OVX, OVX+AMFJ₅, and OVX+AMFJ₁₀ groups were respectively 0.273±0.014 g/cm², 0.263±0.002 g/cm², 0.270±0.010 g/cm², and 0.299±0.012 g/cm² (Fig. 1). As is obvious, BMD of OVX rats was lower than that of SO group (with no statistical significance). *Aronia melanocarpa* fruit juice treatment increased BMD of the treated groups ($F_{2,10}=3.85$, $p=0.05$). Dunnett's multiple comparison test showed a significantly increased BMD in OVX + AMFJ₁₀ group ($p<0.05$ vs. OVX) (Fig. 1).

DISCUSSION

Ovariectomy causes estrogen deficit that mimics the menopause. Typical findings in OVX rats are increased fat accumulation and decreased BMD (8,15).

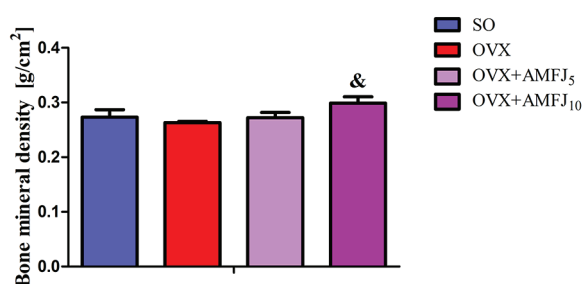


Fig. 1. Femur bone mineral density (BMD, g/cm²); SO—sham-operated rats; OVX—ovariectomized rats; OVX+AMFJ₅—OVX rats treated with AMFJ 5 mL/kg; OVX+AMFJ₁₀—OVX rats treated with AMFJ 10 mL/kg; **p*<0.05 vs. OVX.

In this experiment, OVX rats showed increased fat deposits in comparison with SO animals. This was demonstrated by increased TF and RPF, as well as TF/BW and RPF/BW indices. It might be a result of increased leptin levels leading to leptin insensitivity (16,17). There are data that berries rich in cyanidin-3-glucoside are able to prevent the obesogenic effect of OVX-induced estrogen deficit (18). The *Aronia melanocarpa* fruit juice used in this experiment is very rich in anthocyanins presented by cyanidin-3-glycosides. In several experiments, *Aronia* prevented diet-induced obesity in mice (19,20). In our study, AMFJ did not succeed in antagonizing the effect of ovariectomy on body weight. These results are consistent with the findings of other authors regarding berry fruits (21).

We found that total cholesterol level of OVX animals was insignificantly higher than that of SO rats. This is consistent with other authors' findings (22). In rats estrogen deficit does not always lead to a significant increase in plasma total cholesterol levels. This is probably due to difference in the composition and regulation of the production of plasma lipids, especially of HDL cholesterol (23). *Aronia melanocarpa* fruit juice did not significantly affect cholesterol levels, although there are data that *Aronia* fruits is able to improve lipid metabolism in diet-induced obesity (19,24,25).

In our experiment, a slight, statistically non-significant decrease in BMD was observed in OVX compared to SO rats. This finding is consistent with some other results (8,26). Other experiments performed with Sprague-Dawley rats (27,28) were able

to demonstrate a significant reduction of BMD 3 months after ovariectomy. We found that AMFJ was able to increase BMD in OVX rats, which coincides with the results of Kang et al. (7). This beneficial effect of AMFJ is most probably due to its high polyphenolic content. Polyphenols are found to affect bone metabolism by several mechanisms: inhibition of bone resorption by antioxidant and anti-inflammatory action, stimulation of osteoblastogenesis, and inhibition of osteoclastogenesis, as well as by osteoimmunologic mechanisms (29).

CONCLUSION

Aronia melanocarpa fruit juice was not able to reverse estrogen-deficit-induced obesity in OVX rats. It increased BMD of OVX rats, probably due to its high polyphenolic content.

REFERENCES

- Molina PE. Female Reproductive System. In: Molina PE, editors. Endocrine Physiology. 5th Ed. New York, NY: McGraw-Hill; available from: <http://accessmedicine.mhmedical.com/content.aspx?bookid=2343§ionid=183488853>.
- Lovejoy JC. The menopause and obesity. Prim Care. 2003;30(2):317-25. doi: 10.1016/s0095-4543(03)00012-5.
- Kharode YP, Sharp MC, Bodine PV. Utility of the ovariectomized rat as a model for human osteoporosis in drug discovery. Methods Mol Biol. 2008;455:111-24. doi: 10.1007/978-1-59745-104-8_8.
- Denev PN, Kratchanov CG, Ciz M, Lojek A, Kratchanova MG. Bioavailability and antioxidant activity of black chokeberry (*Aronia melanocarpa*) polyphenols: in vitro and in vivo evidences and possible mechanisms of action. A review. Comp Rev Food Sci Food Safety. 2012;11(5):471-89. doi:10.1111/j.1541-4337.2012.00198.x.
- Shahin L, Phaal SS, Vaidya BN, Brown JE, Joshee N. Aronia (Chokeberry): an underutilized, highly nutraceutical plant. J Med Act Plants. 2019;8(4):46-63. doi: 10.7275/q651-2w57.
- Valcheva-Kuzmanova S, Denev P, Krachanova M, Surleva A, Belcheva A. Composition and antioxidant activity of *Aronia melanocarpa* fruit juice. Varna Med Forum. 2014;3(1):15-20. doi: 10.14748/vmf.v3i1.1109.
- Kang AR, Jung KI, Kim MH. Effects of *Aronia melanocarpa* extracts on menopause symptoms in

- ovariectomized rats. *J Korean Soc Food Sci Nutr*. 2019;47(12):1217-24.
8. Devarreddy L, Hooshmand S, Collins JK, Lucas EA, Chai SC, Arjmandi BH. Blueberry prevents bone loss in ovariectomized rat model of postmenopausal osteoporosis. *J Nutr Biochem*. 2008;19(10):694-9. doi: 10.1016/j.jnutbio.2007.09.004.
 9. Hasona NA, Morsi A, Alghabban AA. The impact of grape proanthocyanidin extract on dexamethasone-induced osteoporosis and electrolyte imbalance. *Comp Clin Pathol*. 2018;27(5):1213-9.
 10. Zhou RP, Lin SJ, Wan WB, Zuo HL, Yao FF, Ruan HB, et al. Chlorogenic acid prevents osteoporosis by Shp2/PI3K/Akt pathway in ovariectomized rats. *PLoS ONE*. 2016;11(12):e0166751. doi: 10.1371/journal.pone.0166751.
 11. Min J, Yuan Z, Zhang Q, Lin S, Wang K, Luo J. Analysis of anti-osteoporosis function of chlorogenic acid by gene microarray profiling in ovariectomy rat model. *Biosci Rep*. 2018;38(4):BSR20180775. doi: 10.1042/BSR20180775.
 12. Oršolić N, Jeleč Z, Nemrava J, Balta V, Gregorović G, Jeleč D. Effect of quercetin on bone mineral status and markers of bone turnover in retinoic acid-induced osteoporosis. *Polish J Food Nutr Sci*. 2018;68(2):149-62.
 13. Singleton VL, Rossi JA. Colorimetry of total phenolics with phosphomolybdic phosphotungstic acid reagents. *Am J Enol Viticult*. 1965;16:144-58.
 14. Howell AB, Reed JD, Krueger CG, Winterbottom R, Cunningham DG, Leahy M. A-type cranberry proanthocyanidins and uropathogenic bacterial anti-adhesion activity. *Phytochemistry*. 2005;66(18):2281-92. doi: 10.1016/j.phytochem.2005.05.022.
 15. Georgieva A, Eftimov M, Todorova M, Kuzmanova V, Kuzmanov A, Kuzmanov K, et al. Effects of ovariectomy-induced estrogen deficit on rat behaviour, lipid metabolism, inflammation, bone mineral density, and turnover. *Folia Med*. 2021;63(3):385-91. doi: 10.3897/folmed.63.e55267.
 16. Meli R, Pacilio M, Raso GM, Esposito E, Coppola A, Nasti A, et al. Estrogen and Raloxifene modulate leptin and its receptor in hypothalamus and adipose tissue from ovariectomized rats. *Endocrinology*. 2004;145(7):3115-21. doi: 10.1210/en.2004-0129.
 17. Sanchez-Mateos S, Alonso-Gonzalez C, Gonzalez A, Martinez-Campa CM, Mediavilla MD, Cos S, et al. Melatonin and estradiol effects on food intake, body weight, and leptin in ovariectomized rats. *Maturitas*. 2007;58(1):91-101. doi: 10.1016/j.maturitas.2007.06.006.
 18. Kaume L, Gilbert WC, Brownmiller C, Howard LR, Devarreddy L. Cyanidin 3-O- β -d-glucoside-rich blackberries modulate hepatic gene expression, and anti-obesity effects in ovariectomized rats. *J Funct Foods*. 2012;4(2):480-8. doi:10.1016/j.jff.2012.02.008.
 19. Park CH, Kim JH, Lee EB, Hur W, Kwon OJ, Park HJ, et al. Aronia melanocarpa extract ameliorates hepatic lipid metabolism through PPAR γ 2 down-regulation. *PLOS ONE*. 2017;12(1):e0169685. doi: 10.1371/journal.pone.0169685.
 20. Yamane T, Kozuka M, Konda D, Nakano Y, Nakagaki T, Ohkubo I, et al. Improvement of blood glucose levels and obesity in mice given aronia juice by inhibition of dipeptidyl peptidase IV and α -glucosidase. *J Nutr Biochem*. 2016;31:106-12. doi: 10.1016/j.jnutbio.2016.02.004.
 21. Elks CM, Terrebonne JD, Ingram DK, Stephens JM. Blueberries improve glucose tolerance without altering body composition in obese postmenopausal mice. *Obesity*. 2015;23(3):573-80. doi: 10.1002/oby.20926.
 22. Liu ML, Xu X, Rang WQ, Li YJ, Song HP. Influence of ovariectomy and 17 β -estradiol treatment on insulin sensitivity, lipid metabolism and post-ischemic cardiac function. *Int J Cardiol*. 2004;97(3):485-93. doi: 10.1016/j.ijcard.2003.11.046.
 23. Lundeen SG, Carver JM, McKean M-L, Winkler RC. Characterization of the ovariectomized rat model for the evaluation of estrogen effects on plasma cholesterol levels. *Endocrinology*. 1997;138(4):1552-8. doi: 10.1210/endo.138.4.5083.
 24. Valcheva-Kuzmanova S, Kuzmanov K, Mihova V, Krasnaliev I, Borisova P, Belcheva A. Antihyperlipidemic effect of Aronia melanocarpa fruit juice in rats fed a high-cholesterol diet. *Plant Foods Hum Nutr*. 2007;62(1):19-24. doi: 10.1007/s11130-006-0036-2.
 25. Zhu Y, Zhang JY, Wei YL, Hao JY, Lei YQ, Zhao WB, et al. The polyphenol-rich extract from chokeberry (*Aronia melanocarpa* L.) modulates gut microbiota and improves lipid metabolism in diet-induced obese rats. *Nutr Metab*. 2020;17:54. doi: 10.1186/s12986-020-00473-9.

26. Jiang Y, Zhao J, Genant HK, Dequeker J, Geusens P. Mineral density and biomechanical properties of spine and femur of ovariectomized rats treated with naproxen. *Bone*. 1998;22(5):509-14. doi: 10.1016/s8756-3282(98)00027-1.
27. Cao DP, Zheng YN, Qin LP, Han T, Zhang H, Rahman K, et al. Curculigo orchoides, a traditional Chinese medicinal plant, prevents bone loss in ovariectomized rats. *Maturitas*. 2008;59(4):373-80. doi: 10.1016/j.maturitas.2008.03.010.
28. Ohta A, Uehara M, Sakai K, Takasaki M, Adlercreutz H, Morohashi T, et al. A combination of dietary fructooligosaccharides and isoflavone conjugates increases femoral bone mineral density and equol production in ovariectomized mice. *J Nutr*. 2002;132(7):2048-54. doi: 10.1093/jn/132.7.2048.
29. Nicolin V, De Tommasi N, Nori SL, Costantinides F, Berton F, Di Lenarda R. Modulatory effects of plant polyphenols on bone remodeling: a prospective view from the bench to bedside. *Front Endocrinol*. 2019;10:494. doi: 10.3389/fendo.2019.00494.